University Animal Care Committee Standard Operating Procedure

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Subject: Hypoxic Studies in Mice
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Location: Queen’s University
Responsibility: Principal Investigators, Research Staff, Veterinary Staff
Purpose: The purpose of this Standard Operating Procedure (SOP) is to describe the method of hypoxic studies in mice.

1. Introduction and Definitions: The purpose of this Standard Operating Procedure (SOP) is to describe the method used inducing the development of pulmonary hypertension by exposing mice to the hypoxic model of disease.

Abbreviations: Animal Care Services ACS, Principal Investigator PI, subcutaneous SC, intravenous IV, intraperitoneal IP, intramuscular IM, per os PO, per rectum PR

2. Materials:
   - ProOx Sensor
     - Calibrated to maintain the level of oxygen at 10% O₂ within the chamber. The alarm should be calibrated to allow for +/- range of 2.0% which, if exceeded, will cause the alarm to sound and the gas to be turned off.
   - CO₂ Sensor
     - Measurements of CO₂, O₂%, temperature, and humidity will be recorded daily.
   - CO₂ Scrub (soda lime -CDMV)
   - Desiccant beads DRIERITE tm- Anhydrous indicating desiccant or equivalent
   - Sterile Cages
   - Water Bottle
   - Nitrogen Gas

3. Procedures:
   - The ProOx sensor will be calibrated and set to maintain an internal oxygen percentage of 10%.
   - Mice will be transferred into a clean caging with adequate food and water.
   - The cage will be placed into the hypoxic chamber with the lids on. If cages are required to be stacked, stacking should be off-set to allow for air circulation around the filter.
   - The gas will be turned on and will fill the chamber until the ProOx sensor reads between 8-12%.
   - The alarm will be turned on to prevent the chamber from deviating from the desired oxygen percentage (+/- 2%).
   - The O₂%, CO₂ level, temperature and humidity will be recorded daily.
   - The mice will be monitored daily by looking into the chamber and assessing overall wellness.
   - The chamber will be opened, and mice will be transferred into a new, clean cage weekly. The gas alarm will be turned off when the chamber is opened.
   - The new cage will be placed back into the chamber, and the gas will be turned on.
• Each time the chamber is opened, the silenced alarm must be reset once the internal O₂% has stabilized.

4. Complications:
• Failure of the sensor may result in a reading of internal O₂% ≤ 8% or ≥ 12%. At this point the alarm will sound and gas will automatically shut off. Research staff will assess the sensor and nitrogen tank and determine the reason for failure before turning the gas back on.
• The scrub will be monitored for a colour change from white to purple as a measure of CO₂ absorption within the chamber. If the scrub is found to indicate high CO₂ absorption (purple colour) this will result in the chamber being opened and the scrub being changed. An additional container of scrub can be added to help maintain the level of CO₂ within the chamber.
• The desiccant will be monitored for a colour change from orange to blue as a measure of internal humidity. Blue desiccant (Drierite or equivalent), will result in the chamber being opened and the desiccant being changed.
• Cages will be monitored to ensure that the mice have adequate food, water and they will also be monitored for water leakage and flooding. For any of these events, the chamber will be opened, and the specific issues resolved.

References: Dr. Archer and Dr. Ormiston Laboratories

Revised: